

Research Article

Open Access

Radon levels in different types of Plants with medicinal properties

Journal of Food Technology

Ali Abid Abojassim^{1*}, HeiyamNajy Hady² and Abdul hussein Abdulameer Kareem³

Madridge

¹University of Kufa/ Faculty of Science/ Physics Department, Al-Najaf-Iraq ^{2,3}University of Kufa/ Education faculty for Girl / Physics Department

Article Info

*Corresponding author: Ali Abid Abojassim Faculty of Science Physics Department University of Kufa Al-Najaf- Iraq E-mail: ali.alhameedawi@uokufa.edu.iq doi: 10.18689/mjft-1000103

Received: August 10, 2016 Accepted: September 1, 2016 Published: November 23, 2016

Citation: Ali Abid Abojassim^{1*}, HeiyamNajy Hady² and Abdul hussein Abdulameer Kareem³. Radon Levels in Different Types of Plants with Medicinal Properties. *Madridge J Food Tech.* 2016; 1(1): 18-21.

Copyright: © 2016 Ali Abid Abojassim et al. This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Published by Madridge Publishers

Abstract

In present study, natural levels of radon-222 in forty selected herbs with medicinal properties present in many stores throughout Iraq were measured to establish any potential radiation hazards. The Solid State Nuclear Track Detectors SSNTDs (CR-39) technique was used to determine these natural levels of radiation. The findings indicate that the radon concentrations ranged from 10.6602.07 Bq/m³ to 53.3034.64 Bq/m³ with an average 26.5373.21Bq/m³. These radon concentration values were lower than those reported in literature. These results show that consumption of the studied plants would impose no health threat to the consumers.

Keywords: Herbs, medicinal plants, radon concentration, nuclear track detector, CR-39.

Introduction

Iraq has a long history and tradition of using herbs and other medicinal plants in traditional therapy which preceded the use of modern drugs and other antibiotics [1-2]. For thousands of years people have used and depended on herbs to treat many diseases. Presently, modern medicine and traditional or alternative medicine [3-5] are not exclusive, rather complementary to each another.

Radon-222 is a noble gas formed from radium (²²⁶Ra) and has a half-life of 3.8 days. Radon-222 can be emitted from the earth, rocks, as well as construction materials, and it can accumulate with its short-lived progeny in the atmosphere within the residences [6]. Exposure and inhalation of ²²²Rn for a short time period may lead to lung cancer. As a result of natural radioactive decay in the soil radon isotope particles, ²²²Rn are released from the soil particles escaping into the atmosphere. The radon release rate from the soil is known as the radon emanation rate or the radon exhalation rate. Radon exhalation is an intricate phenomenon depending on a number of parameters such as soil morphology, radium content in the soil, temperature, atmospheric pressure, soil moisture, rainfall and soil particle size [7]. The Solid State Nuclear Track Detectors or SSNTD's are considered to be most used devices for radon concentration measurements in the ecological fields. They are widely used in detecting and measuring radioactivity in geological samples and studying the influence of pollution in the dwellings [8]. The CR – 39 plastic track detectors were used for the evaluation of radon concentration in different types of herbs used in this study.

Materials and Methods

Sample Collection and Preparation

In September of 2015, forty different samples of medicinal plants were collected from the local markets from various places in Najaf City, Iraq. Samples were classified into groups as shown in Table 1. The cursor in front of each sample represents the sample code, trade name, scientific name, part used and country of origin. The samples were prepared by drying them for two to four days at the temperature from 42 to 44° C to eliminate absorbed moisture and obtain actual, dry weight. The dried samples were ground to a fine powder of equal size particles by using a blender. By using a highly sensitive scale with a tolerance $\pm 0.01\%$, we measured twenty grams of each individual sample for further analysis. Then the samples were placed in containers. Before use, containers were washed with dilute hydrochloric acid and rinsed with distilled water and assigned a code specific to each individual sample.

| No | Sample code | Trade name | Scientific name | Part used | Origin |
|----|----------------|-----------------------------------|----------------------------------|-------------------------|------------------------|
| 1 | H1 | Senna | CassaisennaL. | Leaves | Saudi Arabia |
| 2 | H2 | Safflower | Carthamustinctorius | Flowers | Iran |
| 3 | H3 | Ziziphus | Ziziphusspina-Christi L. | Leaves | Iraq |
| 4 | H4 | Hops | HumuluslupulusL. | Peduncle | Iran |
| 5 | H5 | Peppermint | MenthapiperitaL. | Leaves | Iraq |
| 6 | H6 | Balanite | Balanitesaegyptica(L.) Del. | Fruits | Egypt |
| 7 | H7 | Aelchenan | Anabasis spp. | Leaves | Iraq |
| 8 | H8 | Green tea | Camellia sinensis | Leaves | China |
| 9 | Н9 | Fenugreek | TrigonellafoenumgraecumL. | Seeds | India |
| 10 | H10 | Sweet marjoram | Origanummajorana | Aerial parts | Middle east |
| 11 | H11 | Ginger | Zingiberofficinale | Roots | India |
| 12 | H12 | Greater plantain | Plantago major L. | Peel fruits &seeds | India |
| 13 | H13 | Hawthorn | Crataegus spp. | Leaves | USA |
| 14 | H14 | Chokecherry | PrunusvirginianaL | Seeds | Azerbaijan |
| 15 | H15 | Myrtle | MyrtuscommunisL. | Leaves | Iraq |
| 16 | H16 | White cedar | Thujaoccidentalis | Fruits | Syria |
| 17 | H17 | Rosemary | RosmarinusofficinalisL. | Aerial parts | Mediterra- nean sea |
| 18 | H18 | Chicory | CichoriumintybusL. | Roots, Stalk &leaves | Iraq |
| 19 | H19 | Chamomile | Matricariachamomilla L. | Flowers | Syria |
| 20 | H20 | Sage | Salvia officinalis | Leaves | India |
| 21 | H21 | Maidenhair fern | .Adiantumcapillus -venerisL. | Leaves and Stalk | USA |
| 22 | H22 | Black mustard | Brassica nigra(L.) W.D.J. Koch | Seeds | China |
| 23 | H23 | Cyperus | Cyperusesculentus | Seeds | Egypt |
| 24 | H24 | Hollyhock | Alcearosea L. | Flowers | India |
| 25 | H25 | Ginkgo | Ginkgo biloba. | Seeds | Iran |
| 26 | H26 | Bay leaves | Laurusnobilis | Leaves | Syria |
| 27 | H27 | Corn Mint or Bo He | Menthahalpocalyx | Aerial parts | India |
| 28 | H28 | Black cumin | Nigella sativa L. | Seeds | India |
| 29 | H29 | Roselle | Hibiscus sabdariffaL. | Flowers | Iraq |
| 30 | H30 | Horse tail | EquisetiumarvenseL. | Aerial parts | Egypt |
| 31 | H31 | African Rue | RutachalepensisL. | Aerial parts | Saudi Arabia |
| 32 | H32 | Flax | Linumusitatissimum L. | Seeds | Iran |
| 33 | H33 | Garden Angelica/ Stout bien | Angelica archangelicaL. | Each herb | China |
| 34 | H34 | Yarrow | Achilleamillefolium | Aerial parts | Iran |
| 35 | H35 | Nutgrass | CyperusrotundusL. | Roots and leaves | Saudi Arabia |
| 36 | H36 | Colocynth | Citrulluscolocynthis (L.) Shradc | Fruits | Iraq |
| 37 | H37 | Primrose | Primula vulgaris L. | Flowers | west Asia |
| 38 | H38 | Borage | Boragoofficinales | Flowers | Iran |
| 39 | H39 | Coltsfoot | Tassilagofarfara | Leaves and flowers | North Asia |
| 40 | H40 | Rose of Jericho | Anastaticahierochuntical | Branches | Palestine |

Laboratory Procedure

Measurements were carried out upon 30 days after reaching the radiation equilibrium. Beacon covers were removed rapidly to prevent outside air from entering and changing the atmosphere in the cans. The nuclear detector CR-39 with dimensions of 1cm² and 1mm thick was placed at the middle of the underside of the cover and affixed with an adhesive tape. The edges of the cover were taped and sealed to prevent radon from leaking. The CR-39 detector recorded the presence and effects of alpha particles which resulted from the dissolution of radon gas. The distance between the surfaces of the sample and reagent was 5cm and the sample height was 2cm as shown in Figure 1. We applied the longterm method of 90 day exposure before removing the reagents exposing them to the chemical skimming procedure.



Figure 1: Schematic representation of the plastic container showing the position of the CR-39 detector and the sample tested

Chemical Etching and Microscopic Scanning

Detectors were removed and etched in a 6.25N aqueous solution of NaOH. The detectors were placed inside Pyrex and linked with a wire. The Pyrex was placed inside a water bath. Following the standard protocol of keeping them in the bath at 70 °C for 7 hours, detectors were rinsed with distilled water and allowed to air-dry than placed in a plastic box [9].

The tracks recording the effect of alpha particles at the surface of the CR-39 nuclear detectors were observed by using novel optical microscope at 400x magnification as shown in. Figure 2. A total of five optic view fields were selected for taking the readings for each individual sample.



Figure 2: The effects of alpha particles chemical etching as seen at 400x by optical microscope

Calculation of Radon concentrations

The density of the tracks ρ in the detectors was calculated according to the following equation [10].

 Table 1: List of herbs used in the study

$$\rho = \frac{N_{average}}{A}\dots\dots\dots\dots(1)$$

Where, ρ is track density (Track /cm²), Nis an average of total tracks (Track) and A is an area of a view field (cm²).

Radon concentration (C_{Rn}) in Bq/m³ unit are calculated by the following equations [11-12].

Where, k : is the calibration factor in terms of $(track.cm^{-2}/Bq.d.m^{-3})$ which is the same value as reported in many works [12-15]. T is the exposure time (d).

The value for radon activity (A_{Rn}) and specific radon activity $(S.A_{Rn})$ can be found based on radon concentrations, volume of container (V) and mass of a sample (m) as it follows: [8][16][17]

$$A_{Rn} = C_{Rn} V \dots \dots \dots \dots \dots (3)$$
$$S. A_{Rn} = \frac{A_{Rn}}{m} \dots \dots \dots \dots \dots (4)$$

Results and Discussion

Table 2 presents radon concentration and specific activity for medicinal plant samples originated in different countries but obtained at the local markets. Our study shows that the lowest value of radon concentration was found in H6 Egypt sample, which was (10.6602.07 Bq/m3), while the highest average value was found in H35 Saudi Arabia sample, having values of (53.3034.64 Bg/m³) as shown in Figure 3. The same sample from Saudi Arabia had shown specific radon activity. The specific radon activity varied from 0.003±0.0003Bg/kg in H₂O samples to 0.7790.085 Bq/kg in H30 samples with an average 0.2830.035Bq/kg. These results indicate that the reasons for variation of radon concentration in different medicinal plant samples are the result of chemical reaction by which radon transfer occurs from the soil solids into the water solution within the soil becoming absorbed by the roots and translocated throughout the plants. The uptake of these radio nuclides from the soil solution is controlled by plant physiology. Our findings are similar and very close to the reported results cited in the literature as presented in Table 3.

| Sample code | C_{a} (Bg/m ³) | SA, (Ba/ka) | |
|-------------|------------------------------|----------------------------|--|
| Sample code | | 5.7 _{Rn} (Dq/ Kg) | |
| H1 | 50.6384.52 | 0.4700.042 | |
| H2 | 39.9774.01 | 0.6490.085 | |
| H3 | 31.9823.59 | 0.0310.277 | |
| H4 | 21.3212.93 | 0.0290.213 | |
| H5 | 15.9912.54 | 0.0300.188 | |
| H6 | 10.6602.07 | 0.0080.046 | |
| H7 | 29.3163.44 | 0.0230.200 | |
| H8 | 18.6562.74 | 0.0230.161 | |
| H9 | 23.9863.11 | 0.0220.173 | |
| H10 | 13.3252.32 | 0.0270.157 | |
| H11 | 11.7262.17 | 0.0170.095 | |
| H12 | 19.9882.84 | 0.0280.199 | |
| H13 | 22.6533.02 | 0.0490.368 | |
| H14 | 20.5212.87 | 0.0240.177 | |
| H15 | 17.3232.64 | 0.0260.173 | |
| H16 | 23.3203.06 | 0.0210.159 | |

| | | * |
|---------|------------|-------------|
| H17 | 16.7902.60 | 0.0330.218 |
| H18 | 19.4552.80 | 0.0600.421 |
| H19 | 25.3193.19 | 0.0590.470 |
| H20 | 21.9872.98 | 0.00030.003 |
| H21 | 22.9203.04 | 0.0490.372 |
| H22 | 23.7203.09 | 0.0260.205 |
| H23 | 34.6473.74 | 0.0280.264 |
| H24 | 45.3074.27 | 0.0460.490 |
| H25 | 27.9843.36 | 0.0310.259 |
| H26 | 35.9793.81 | 0.0300.292 |
| H27 | 21.0542.91 | 0.0340.248 |
| H28 | 17.9892.69 | 0.0200.137 |
| H29 | 12.7922.27 | 0.0130.075 |
| H30 | 47.9734.40 | 0.0710.779 |
| H31 | 18.3892.72 | 0.0350.239 |
| H32 | 31.3153.55 | 0.0270.239 |
| H33 | 22.1202.98 | 0.0350.261 |
| H34 | 28.6503.40 | 0.0550.465 |
| H35 | 53.3034.64 | 0.0240.277 |
| H36 | 25.9853.24 | 0.0280.225 |
| H37 | 23.4533.07 | 0.1000.762 |
| H38 | 17.5902.66 | 0.0430.285 |
| H39 | 27.7173.34 | 0.0620.514 |
| H40 | 19.7222.82 | 0.0200.142 |
| Max | 53.3034.64 | 0.7790.085 |
| Min | 10.6602.07 | 0.00030.003 |
| Average | 26.5373.21 | 0.2830.035 |
| | | |

 Table 2: The radon concentration and specific radon activity in medicinal plants

| Country | Average of radon concentrations (Bq/m ³) | Reference | |
|--------------|--|-----------|--|
| Iraq(Bagdad) | 19.2 | [8] | |
| Morocco | 10.2 | [18] | |
| Iraq(Najaf) | 26.5 | This work | |

 Table 3: Our results showing radon concentration compared to findings cited in literature



Figure 3: Radon concentration in medicinal plants

Conclusion

The forty herb samples collected from the local markets from various places in Iraq were evaluated for any potential radiation hazards by using SSNTDs (CR-39 detector). Our research indicates that the highest concentration of radon is found in roots and leaves, while the lowest radon concentration value is found in fruits. The radiological study shows that consumption of the studied medicinal plants would impose no health threat to the consumers.

References

- 1. Rukangira E. The African Herbal Industry: Constraints and Challenges. [(Accessed on 26 March 2014)]. (2012).
- Kabir OA, Olukayode O, Chidi EO, Christopher CI, Kehinde AF. Screening of crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine for anti-methicillin resistant Staphylococcus aureus activity. *BMC complementary and alternative medicine*. 2005; 5(1): doi: 10.1186/1472-6882-5-6
- Priya KS, Gnanamani A, Radhakrishnan N, Babu M. Healing potential of Datura alba on burn woundsin albino rats. *Journal of ethno pharmacology*. 2002; 83(3): 193–199.
- Steenkamp V, Mathivha E, Gouws MC, Rensburg CEJ. Studies on antibacterial, antioxidant and fibroblast growth stimulation of wound healing remedies from South Africa. *Journal of ethnopharmacology*. 2004; 95(2-3): 353-357. doi: 10.1016/j.jep.2004.08.020
- Patel Mrudula, Maeve MC. Antifungal activity of the plant Dodonaeaviscose var. angustifolia on Candida albicansfrom HIVinfected patients. *Journal of ethnopharmacology*. 2008; 118(1): 173-176. doi: 10.1016/j.jep.2008.03.009
- Rawat A, Jojo PJ, Khan AJ, Tyagi RK, Rajendra P. Radon exhalation rate in building materials. *International Journal of Radiation Applications and Instrumentation. Part D. Nuclear Tracks and Radiation Measurements.* 1991; 19(1-4): 391-394. doi: 10.1016/1359-0189(91)90223-5
- Khan MS, Srivastava DS, Azam A. Study of radium content and radon exhalation rates in soil samples of northern India. *Environmental Earth Sciences*. 2012; 67(5): 1363-1371. doi: 10.1007/s12665-012-1581-7
- 8. Dawser HG. Measurement of Radon Concentration in Henna Plant Using Etched Track Detectors. *Al- Mustansiriya J. Sci.* 2011; 22(1): 139- 144.
- 9. Mohammad AM, Ahamed IK, Ahmed YS. Measurement of Uranium Concentration in Some Soil Samples in Jalawla'a City Using CR-39 Detector. *Journal of Al-Nahrain university*. 2013; 16(1): 112-116.

- 10. Ali AA, Hamidawi A, Afnan AH. Radiation Hazards Due to Radon Concentrations in Dwellingsof Kufa Technical Institute, 'Iraq. *Physics International.* 2016; 7(1): 28-34. doi: 10.3844/pisp.2016.28.34
- 11. Al-Kofahi M, Khader B, Lehlooh A, Kullab M, Abumurad K, Al- Bataina B, Nucl., Tracks Radiat. *Meas.* 1992; 20: 377-382.
- 12. Rasas M. Measurement of Radon and Its Daughter's Concentrations In Indoor and Outdoor Throughout Gaza Strip. *Journal of the Association of Arab Universities for Basic and Applied Sciences.* 2003; 11(1): 21-26. doi: 10.1016/j.jaubas.2011.10.003
- AL-Kofahl MM, Khader BR, Lehlooh AD, Kullar MK. Measurement of Radon-222 in Jordanian Dwellings. International Journal of Radiation Applications and Instrumentation. Part D. Nuclear Tracks and Radiation Measurements. 1991; 20(2): 377-82. doi: 10.1016/1359-0189(92)90068-7
- 14. Mayya YS, Eappen KP, Nambi KSV. Radiat. Prot. Dosim, 1998; 77(3): 177-184.
- 15. M. Sersawi, Study of the chronic radiation exposure situation in Gaza.(M. Sc. Thesis) Islamic University of Gaza in Physics, 2007.
- HeshamYousef A, GehadSaleh M, El-Farrash AH, Hamza A. Radon exhalation rate for phosphate rocks samples using alpha track detectors. *Journal of Radiation Research and Applied Sciences*. 2016; 9(1): 41-46. doi: 10.1016/j.jrras.2015.09.002
- 17. Tykva R, Jozef S. Low-level environmental radioactivity: sources and evaluation. U.S.A, 1995.
- Oufni L, Manaut N, Taj S, Manaut B. Determination of Radon and Thoron Concentrations in Different Parts of Some Plants Used in Traditional Medicine Using Nuclear Track Detectors. *American Journal of Environmental Protection*. 2013; 1(2): 34-40.